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Introduction

The E.Z.N.A.™ family of products is an innovative system that radically simplifies the extraction and purification of nucleic acids from a variety of sources. The key to this system is the new HiBind® matrix that specifically, but reversibly, binds DNA or RNA under certain optimal conditions allowing proteins and other contaminants to be removed. Nucleic acids are easily eluted with deionized water or a low salt buffer.

The E.Z.N.A.™ Plasmid Mini Kit combines the power of HiBind® technology with the time tested consistency of alkaline-SDS lysis of bacterial cells to deliver high quality DNA under 20 minutes. Omega Bio-Tek's (OBI) mini-columns facilitate the binding, washing, and elution steps thus enabling multiple samples to be simultaneously processed. Yields vary according to plasmid copy number, E.coli strain, and conditions of growth; 1.5 ml of overnight cultures in LB medium typically produce 8-12 µg of plasmid DNA. OBI's E.Z.N.A.™ Plasmid Mini Kit II, is upscaled for the isolation of low copy-number plasmids and yields 40-75 µg of DNA from 10-15 ml cultures when using high copy plasmids. OBI's E.Z.N.A.™ Plasmid DNA Systems are suitable for automated fluorescent DNA sequencing, restriction endonuclease digestion, and other manipulations.

Benefits of the E.Z.N.A.™ Plasmid Mini Kits

- **Fast -** Plasmid DNA recovery under 20 minutes
- **Reliability -** with optimized buffers that guarantee pure DNA
- **Safety -** due to no organic extractions
- **Quality -** ensures that purified DNA will be suitable for any application

Storage and Stability

All E.Z.N.A.™ Plasmid Mini Kit components are guaranteed for at least 12 months from the date of purchase when stored as follows: Solution I/ RNase A mixture at 4°C, and all other materials at 22-25°C.

Kit Contents

E.Z.N.A.™ Plasmid Mini Kit I

Product Number	D6942-00 D6943-00 D6944-00	D6942-01 D6943-01 D6944-01	D6942-02 D6943-02 D6944-02
Purification Times	5 Preps	100 Preps	200 Preps
HiBind® Miniprep Columns (I)	5	100	200
2 ml Collection Tubes	5	100	200
Solution I *	3 ml	30 ml	60 ml
Solution II	3 ml	30 ml	60 ml
Solution III	3 ml	40 ml	80 ml
Buffer HB	3 ml	60 ml	120 ml
Elution Buffer	-	20 ml	40 ml
DNA Wash Buffer Concentrate	2 ml	2 x 20 ml	3 x 20 ml
RNase A	pre-added	200 µl	400 µl
Instruction Booklet	1	1	1

E.Z.N.A.™ Plasmid Mini Kit II

Product Number	D6945-00	D6945-01	D6945-02
Purification Times	5 preps	50 preps	200 preps
HiBind® Miniprep Columns (II)	5	50	200
2 ml Collection Tubes	5	50	200
Solution I *	5 ml	30 ml	120 ml
Solution II	5 ml	30 ml	120 ml
Solution III	5 ml	40 ml	2 x 80 ml
Buffer HB	5 ml	30 ml	120 ml
Elution Buffer	-	20 ml	40 ml
DNA Wash Buffer Concentrate	2 ml	20 ml	3 x 20 ml
RNase A	pre-added	100 µl	400 µl
Instruction Booklet	1	1	1

*** RNase A has been added for you to Solution I in Trial Kit D6942-00, D6943-00, D6944 & D6945-00**

Before Starting

It is strongly advised that you familiarize yourself with the entire booklet before starting. E.Z.N.A ™ Kits are designed to be simple, fast, and reliable provided that all steps are followed diligently. Prepare all components, and have the necessary materials as outlined before starting.

Materials Supplied by User

- Microcentrifuge capable of at least 13,000 x g
- Sterile 1.5 ml or 2 ml Centrifuge Tubes
- Absolute (96%-100%) Ethanol
- Sterile 15-50 ml centrifuge tubes (Product No. D6945 only)

Important Notes

- **Add vial of RNase A provided to Solution I** (for 50 prep and 200 prep Kits). Store at 4°C.
- Check Solution II and Solution III for salt precipitation before use. Redissolve any precipitation by warming to 37°C.
-

Dilute DNA Wash Buffer Concentrate with absolute ethanol (~96-100%) as follows and Store at room temperature.	
D6942/D6943/D6944/D6945-00	Add 8 ml of absolute ethanol
D6942/D6943/D6944/D6945-01	Add 80 ml of absolute ethanol/bottle
D6942/D6943/D6944/D6945-02	Add 80 ml of absolute ethanol/bottle

- All steps must be carried out at room temperature.

Safety Information

When working with chemicals, always wear a suitable lab coat, disposable gloves, and protective goggles. The following risk and safety phrases apply to components of the E.Z.N.A.™ Plasmid Kits.

- **Solution II** contains sodium hydroxide: irritant. *R36/38,S13-26-36-46
- **Solution III** contains acetic acid: irritant. *R36/38,S13-26-36-46
- **Buffer HB** contains isopropanol: flammable. *R10
- **RNase A** contains ribonuclease: sensitizer. *R42/43, S23-24-26-36/37

E.Z.N.A.™ Plasmid Mini Protocol I (Product No. D6942, D6943 & D6944)

1. **Isolate a single colony from a freshly streaked selective plate, and inoculate a culture of 1- 5 ml LB medium containing the appropriate selective antibiotic. Incubate for ~ 12-16 hr at 37°C with vigorous shaking (~ 300 rpm)**. Use a 10-20 ml culture tube or a flask with a volume of at least 4 times the volume of the culture. It is strongly recommended that an endA negative strain of *E.coli* be used for routine plasmid isolation. Examples of such strains include DH5α® and JM109®.

2. **Pellet 1.5-5.0 mL bacteria by centrifugation at 10,000 x g for 1 min at room temperature.** Decant or aspirate medium and discard.

3. **Resuspend the bacterial pellet by adding 250 µl of Solution I/RNase A and vortexing (or pipetting up and down).** Complete resuspension (no visible cell clumps) of cell pellet is vital for obtaining good yields.

4. **Add 250 µl of Solution II and gently mix by inverting and rotating the tube several times to obtain a clear lysate. A 2 minutes incubation may be necessary.** Avoid vigorous mixing as this will shear chromosomal DNA and lower plasmid purity. Do not allow the lysis reaction to proceed more than 5 min. (Store Solution II tightly capped when not in use to avoid acidification from CO_2 in the air.)

5. **Add 350 µl of Solution III and mix immediately by inverting the tube several times until a flocculent white precipitate forms.** It is vital that the solution is mixed thoroughly, and immediately after the addition of Solution III to avoid localized precipitation.

6. **Centrifuge at ≥10,000 x g for 10 minutes at room temperature.** A compact white pellet will form. Promptly proceed to the next step.

7. **Add the cleared supernatant by CAREFULLY aspirating it into a clean HiBind® Miniprep Column (I) assembled in a provided 2 ml collection tube**. Ensure that the pellet is not disturbed and that no cellular debris has been carried over into the column. **Centrifuge for 1 min at 10,000 x g at room temperature to completely pass lysate through the HiBind® Miniprep Column (I).**

8. **Discard flow-through liquid and re-use the 2 ml collection tube. Add 500 µl of Buffer HB to wash the HiBind® Miniprep Column (I). Centrifuge for 1 min at 10,000 x g at room temperature to completely pass solution through the HiBind® Miniprep Column (I).**This step ensures that residual protein contaminations are removed, thus ensuring high quality DNA that will be suitable for downstream applications.

9. **Discard flow-through liquid and re-use the 2 ml collection tube. Add 700 µl of DNA Wash Buffer diluted with absolute ethanol to wash the HiBind® Miniprep Column (I). Centrifuge for 1 min at 10,000 x g at room temperature to completely pass solution through the HiBind® Miniprep Column (I) and discard flow-through liquid.**

   Note: DNA Wash Buffer Concentrate must be diluted with absolute ethanol before use. See label for directions. If refrigerated, DNA Wash Buffer must be brought to room temperature before use.

10. **Optional Step:** Repeat wash step with another 700 µl of DNA Wash Buffer diluted with absolute ethanol.

11. **Centrifuge the empty column for 2 min at ≥13,000 x g to dry the column matrix.** Do not skip this step, it is critical for good yields.

12. **Place the column into a clean 1.5 ml microcentrifuge tube. Add 30 µl to 50 µl**(Depending on desired concentration of final product) **of Elution Buffer** (10mM Tris-HCl, pH 8.5) **or sterile deionized water directly onto the column matrix and let it sit at room temperature for 1-2 minutes. Centrifuge for 1 min at ≥ 13,000 x g to elute DNA.** An optional second elution will yield any residual DNA, though at a lower concentration.

13. **Yield and quality of DNA**: **Determine the absorbance of an appropriate dilution of the sample at 260 nm and then at 280 nm.** The DNA concentration is calculated as follows:

    DNA concentration = A_{260} x 50 x (Dilution Factor) µg/ml

    A ratio of (A_{260}) / (A_{280}) is an indication of nucleic acid purity. A value greater than 1.8 indicates > 90% nucleic acid purity. Alternatively, yield (as well as quality) can sometimes be best determined by agarose gel/ethidium bromide electrophoresis by comparison to DNA samples of known concentrations. Typically, the majority of the eluted DNA is in monomeric supercoil form, although concatamers may also be present.

Vacuum / Spin Protocol for Plasmid Extraction

(Product No. D6943 only, V-Spin Column)

Carry out bacterial culture, lysis and neutralization as indicated in steps 1-6 of the previous protocol. Instead of continuing with centrifugation, follow the steps outlined below.

For your safety wear safety glasses when working near a manifold under pressure.

1. Prepare the vacuum manifold according to manufacturer's instructions, and connect the V-Spin HiBind® Miniprep Column (I) to the manifold.

2. **Load the clear supernatant from step 6 onto the V-Spin HiBind® Miniprep Column (I).**

3. Switch on the vacuum source to draw the sample through the column. Turn off the vacuum.

4. **Add 500 µl of Buffer HB to wash the HiBind® Miniprep Column (I).** Turn on the vacuum source to completely draw the solution through the HiBind® Miniprep Column (I).

5. **Add 700 µl of DNA Wash Buffer diluted with absolute ethanol to wash the HiBind® Miniprep Column (I).** Turn on the vacuum source to completely draw the DNA Wash Buffer through the HiBind® Miniprep Column (I). **Repeat this step with another 700 µl of DNA Wash Buffer.**

   Note: DNA Wash Buffer Concentrate must be diluted with absolute ethanol before use. See label for directions. If refrigerated, DNA Wash Buffer must be brought to room temperature before use.

6. **Assemble the HiBind® Miniprep Column (I) into a provided 2 ml collection tube and transfer the column into a microcentrifuge. Spin at ≥ 13,000 x g for 2 minute to dry the column.**

7. **Place the column into a clean 1.5 ml microcentrifuge tube and add 30-50 µl of Elution Buffer (10 mM Tris-HCl, pH 8.5) or deionized water.** Let it stand for 1-2 minutes and centrifuge for 1 minute at ≥13,000 x g to elute DNA. (You may proceed to step 13 of the previous protocol for obtaining the yield and quality of your purified Plasmid DNA.)

Low-Copy Number Plasmids

Low copy number plasmids generally give 0.1-0.5 µg of DNA per ml in overnight cultures. For routine screening of recombinant clones, a 5 ml culture should provide ample material for agarose gel visualization or restriction digest analysis. However, the method can be modified to essentially double the yield if necessary. Start with a 10 ml bacterial culture, and pellet cells either successively (1.5 ml of culture at a time), or by centrifuging for 10 min at 5,000 x g in a 15 ml centrifuge tube. Proceed to step 3 (page 5) and double the volumes of Solutions I, II, and III. Continue as above using only one HiBind® Miniprep Column (I) per 10 ml culture. There is no need to increase the volumes of Buffer HB and DNA Wash Buffer used.

Note: This method is not recommended for high-copy number plasmids, due to the HiBind® Mini Column (I) becoming quickly saturated. In this situation we recommend the processing of multiple samples from the same culture. Alternatively, use the E.Z.N.A.™ Plasmid Mini Kit II (product No. D6945), a member of the E.Z.N.A.™ Nucleic Acid Purification family that allows processing of 10-15 ml cultures using the mini-column format, and generally yielding 40-70 µg of plasmid DNA from high-copy number plasmids.

E.Z.N.A.™ Plasmid Mini Protocol II (Product No. D6945)

Note: Using the following protocol with the HiBind® Miniprep Columns (I) from E.Z.N.A.™ Plasmid Miniprep Kit I (D6942/D6943/D6944) will not significantly improve yields due to the lower column binding capacity. The E.Z.N.A.™ Plasmid Mini Kit II allows rapid and reliable isolation of greater than 50 µg plasmid DNA using the spin-column format.

Procedure

1. **Isolate a single colony from a freshly streaked selective plate, and inoculate a culture of 10-15 ml LB medium (50 µg/ml) containing the appropriate selective antibiotic. Incubate for ~ 12-16 hr at 37°C with vigorous shaking (~ 300 rpm)**. Use a flask or tube with a volume of at least 4 times the volume of the culture. It is strongly recommended that an endA negative strain of *E.coli* be used for routine plasmid isolation. Examples of such strains include DH5α® and JM109®.

2. **Pellet 10-15 ml bacteria by centrifugation at 5,000 x g for 10 min at room temperature,** preferably in a swinging bucket rotor. Decant or aspirate medium and discard.

3. **Resuspend the bacterial pellet by adding 500 µl of Solution I/RNase A and vortexing (or pipetting up and down).** Complete resuspension of cell pellet is vital for obtaining good yields.

4. **Transfer the cell suspension into a 2 ml microfuge tube and add 500 µl of Solution II. Gently mix by inverting and rotating the tube several times to obtain a clear lysate.** A 2-5 min incubation at room temperature may be necessary. Avoid vigorous mixing as this will shear chromosomal DNA and lower plasmid purity. Do not allow the lysis reaction to proceed more than 5 min. (Store Solution II tightly capped when not in use to avoid acidification of Solution II from CO_2 in the air.)

5. **Add 700 µl of Solution III and mix immediately by inverting the tube several times until a flocculent white precipitate forms.** It is vital that the solution is mixed thoroughly, and immediately after the addition of Solution III to avoid localized precipitation.

6. **Centrifuge at ≥10,000 x g for 10 minutes at room temperature.** A compact white pellet will form. Promptly proceed to the next step.

7. **Add the cleared supernatant by CAREFULLY aspirating it into a clean HiBind® Miniprep Column (II) assembled in a provided 2 ml collection tube**. Ensure that the pellet is not disturbed and that no cellular debris has been carried over into the column. Centrifuge for 1 min at 10,000 x g at room temperature to completely pass lysate through the HiBind® Miniprep Column (II). Discard flow-through liquid and add the remaining lysate to the column and centrifuge as above.

8. **Discard flow-through liquid and re-use the collection tube. Add 500 µl of Buffer HB to wash the HiBind® Miniprep Column (II). Centrifuge for 1 min at 10,000 x g at room temperature to completely pass solution through the HiBind® Miniprep Column (II).** This step ensures that residual protein contaminations are removed, thus ensuring high quality DNA that will be suitable for downstream applications.

9. **Discard flow-through liquid and add 700 µl of DNA Wash Buffer diluted with absolute ethanol to wash the HiBind® Miniprep Column (II). Centrifuge for 1 min at 10,000 x g at room temperature to completely pass solution through the HiBind® Miniprep Column (II), and discard flow-through liquid.**

   Note: DNA Wash Buffer Concentrate must be diluted with absolute ethanol before use. See label for directions. If refrigerated, DNA Wash Buffer must be brought to room temperature before use.

10. **Optional Step: Repeat wash step with another 700 µl of DNA Wash Buffer diluted with absolute ethanol.**

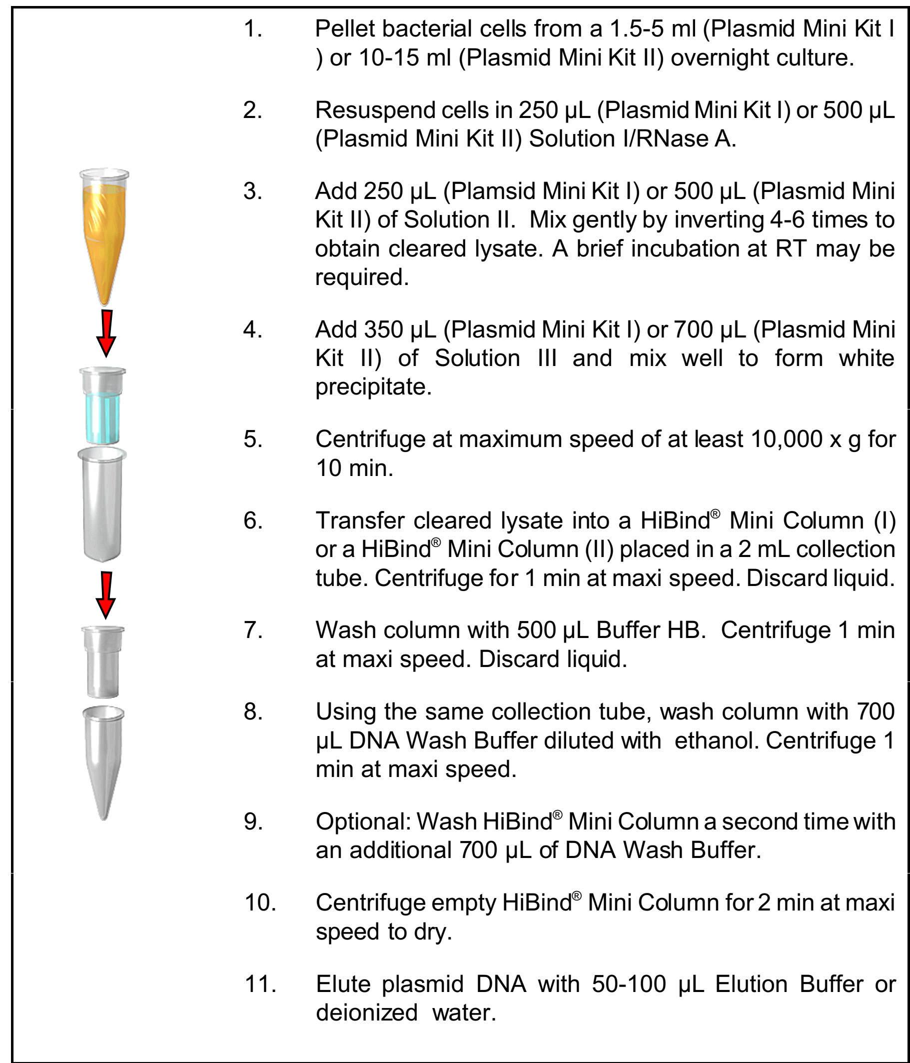
11. **Centrifuge the empty column for 2-3 min at ≥ 13,000 x g to dry the column matrix.** Do not skip this step, it is critical for good yields.

12. **Place the column into a clean 1.5 ml microcentrifuge tube. Add 80 µl to 100 µl** (Depending on desired concentration of final product) **of Elution Buffer** (10mM Tris-HCl, pH 8.5) **or sterile deionized water directly onto the column matrix and let it sit at room temperature for 1-2 minutes. Centrifuge for 1 min at ≥ 13,000 x g to elute DNA.** An optional second elution will yield any residual DNA, though at a lower concentration.

13. **Yield and quality of DNA**: **Determine the absorbance of an appropriate dilution of the sample at 260 nm and then at 280 nm.**
    The DNA concentration is calculated as follows:
    DNA concentration = A_{260} x 50 x (Dilution Factor) µg/ml

Short Mini Kits Protocol for Experienced Users

Note: All steps are to be performed at room temperature. Refer to page 4 for important notes on the preparation of components.



1. Pellet bacterial cells from a 1.5-5 ml (Plasmid Mini Kit I) or 10-15 ml (Plasmid Mini Kit II) overnight culture.
2. Resuspend cells in 250 µL (Plasmid Mini Kit I) or 500 µL (Plasmid Mini Kit II) Solution I/RNase A.
3. Add 250 µL (Plamsid Mini Kit I) or 500 µL (Plasmid Mini Kit II) of Solution II. Mix gently by inverting 4-6 times to obtain cleared lysate. A brief incubation at RT may be required.
4. Add 350 µL (Plasmid Mini Kit I) or 700 µL (Plasmid Mini Kit II) of Solution III and mix well to form white precipitate.
5. Centrifuge at maximum speed of at least 10,000 x g for 10 min.
6. Transfer cleared lysate into a HiBind® Mini Column (I) or a HiBind® Mini Column (II) placed in a 2 mL collection tube. Centrifuge for 1 min at maxi speed. Discard liquid.
7. Wash column with 500 µL Buffer HB. Centrifuge 1 min at maxi speed. Discard liquid.
8. Using the same collection tube, wash column with 700 µL DNA Wash Buffer diluted with ethanol. Centrifuge 1 min at maxi speed.
9. Optional: Wash HiBind® Mini Column a second time with an additional 700 µL of DNA Wash Buffer.
10. Centrifuge empty HiBind® Mini Column for 2 min at maxi speed to dry.
11. Elute plasmid DNA with 50-100 µL Elution Buffer or deionized water.

Troubleshooting Guide

Please use this guide to solve any problems that may arise. We hope that it will aid in clearing up any questions for you, if for any reason you need further assistance the scientists at Omega Bio-Tek, Inc. are always happy to answer any questions you may have about either the information and procedures of this manual or molecular biology applications (see page 16 for contact information).

Possible Problems and Suggestions

Low DNA yields

Problem	Suggestion
a) Poor Cell Lysis	Only use LB or YT medium containing antibiotic. Do not use more than 5 ml (with high copy number plasmids or 10 ml with low copy number plasmids) culture with the basic protocol I. Do not use more than 15 ml culture with the protocol II.
	Cells may not have been dispersed adequately prior to the addition of Solution II. Make sure to vortex cell suspension to completely disperse.
	Increase incubation time with Solution II to obtain a clear lysate.
	Solution II if not tightly closed, may need to be replaced. Prepare as follows: 0.2 N NaOH, 1% SDS.
b) Bacterial Clone is overgrown or not fresh.	Do not incubate cultures for more than 16 hr at 37°C. Storage of cultures for extended periods prior to plasmid isolation is detrimental.
c) Low elution efficiency	The pH of Elution Buffer or water must be ≥8.0

Possible Problems and Suggestions

d) Low copy-number plasmid used	Such plasmids may yield as little as 0.5µg of DNA from a 5 ml overnight culture. Increase culture volume to 10 ml and follow suggested modifications with Plasmid Miniprep Kit I or use the Plasmid Miniprep Kit II (25 ml culture)

No DNA Eluted

e) DNA Wash Buffer Concentrate not diluted with 96-100% ETOH.	Prepare DNA Wash Buffer Concentrate according to instructions on page 4.

High molecular weight DNA contamination of product.

f) Over mixing of cell lysate upon addition of Solution II.	Do not vortex or mix aggressively after adding Solution II.
g) Culture overgrown	Overgrown culture contains lysed cells and degraded DNA. Do not grow cell for longer than 16 hours.

RNA visible on agarose gel

h) RNase A not added to Solution I.	Check that RNase A provided with the kit has been used. If Solution I is more than 6 months old, add more RNase A.

Plasmid DNA floats out of well while loading agarose gel

j) Ethanol has not completely been removed from column following wash steps.	Centrifuge column as instructed to dry the column before elution

Ordering Information

Omega Bio-Tek, Inc.'s line of E.Z.N.A.™ Plasmid Isolation Systems

Product	Applications	Cat. No.
Standard E.Z.N.A.™ Plasmid Isolation System		
E.Z.N.A.™ Plasmid Mini Kit I	Isolation of up to 30 µg Plasmid DNA	D6942-01/02 D6943-01/02
E.Z.N.A.™ Plasmid Mini Kit II	Isolation of up to 70 µg Plasmid DNA	D6945-01/02
E.Z.N.A.™ Plasmid Midi Kit	Isolation of up to 250 µg Plasmid DNA	D6904-03/04
E.Z.N.A.™ Plasmid Maxip Kit	Isolation of up to 1.5 mg Plasmid DNA	D6922-01/02
E.Z.N.A.™ Fastfilter® Plasmid Midi Kit	Isolation of up to 250 µg Plasmid DNA, featuring filter syringes for lysate clearance	D6905-03/04
E.Z.N.A.™ Fastfilter® Plasmid Maxi Kit	Isolation of up to 1.5 mg Plasmid DNA, featuring filter syringes for lysate clearance	D6924-01/03/04
E-Z 96® Fastfilter® Plasmid Isolation Kit	Isolation of Plasmid DNA using a 96-well format	D1097-01/02
E-Z 96® SE Plasmid Isolation Kit	Isolation of plasmid DNA using a single plate	D1095-01/02
E.Z.N.A.™ Yeast Plasmid Isolation Kit	Isolation fo Yeast Plasmid DNA	D3476-01/02 D3376-01/02
E.Z.N.A.™ Endotoxin Free Plasmid Isolation System		
E.Z.N.A.™ Endo-Free Plasmid Miniprep Kit I	Isolation of up to 30 µg Endotoxin free Plasmid DNA	D6948-01/02
E.Z.N.A.™ Endo-Free Plasmid Miniprep Kit II	Isolation of up to 70 µg Endotoxin free Plasmid DNA	D6950-01/02

E.Z.N.A.™ Endo-Free® Plasmid Midiprep Kit	Isolation of up to 250 µg Endotoxin free Plasmid DNA, featuring filter syringes for lysate clearance	D6915-01/03/04
E.Z.N.A.™ Endo-Free® Plasmid Maxiprep Kit	Isolation of up to 1.5 mg Endotoxin free Plasmid DNA, featuring filter syringes for lysate clearance	D6926-01/03/04
E.Z.N.A.™ High Performance Plasmid Isolation System		
E.Z.N.A.™ HP Plasmid Miniprep Kit I	Isolation of up to 30 µg of High Purity Plasmid DNA	D7042-01/02 D7043-01-02
E.Z.N.A.™ HP Plasmid Miniprep Kit II	Isolation of up to 70 µg of High Purity Plasmid DNA	D7045-01-02
E.Z.N.A.™ HP Plasmid Midiprep Kit	Isolation of up to 200 µg of High Purity Plasmid DNA	D7004-01/02
E.Z.N.A.™ HP Plasmid Maxiprep Kit	Isolation of up to 1.5 mg of High Purity Plasmid DNA	D7022-01/02
E.Z.N.A.™ Single Strand Phage DNA Isolation Kits		
E.Z.N.A.™ M13 Isolation Kit	Isolation of up to 15µg of single stranded phage DNA	D6900-01/02
E-Z 96® M13 Isolation Kit	Isolation of up to 15µg of single stranded phage DNA using a 96-well format	D1900-01
E.Z.N.A.™ Large Construct DNA Isolation Kits		
E.Z.N.A.™ BAC/PAC DNA Isolation Kit	Effective purification of BAC or PAC DNA	D2156-01/02
E-Z 96® BAC/PAC DNA Isolation Kit	Parallel purification of BAC or PAC DNA using a 96-well format	D1056-01/02
E-Z 96 Fastfilter BAC/PAC DNA Kit	Parallel purification of BAC or PAC DNA using a 96-well format	D1055-01/02
E.Z.N.A.™ Magnetic Beads Plasmid Isolation Kits		
E-Z 96® Mag-Bind® Plasmid Isolation Kit	Isolation of Plasmid DNA using a 96-well format and paramagnetic beads	M1256-01/02
E-Z 96® Mag-Bind® Endo-Free® Plasmid Isolation Kit	Isolation of Endotoxin free Plasmid DNA using a 96-well format and paramagnetic beads	M1258-01/02
Mag-Bind® Plasmid Maxiprep Kit	Isolation of up to 1.5 mg Plasmid DNA using magnetic beads technology	M1257-01/02
Mag-Bind® Plasmid Megaprep Kit	Isolation of up to 5 mg Plasmid DNA using magnetic beads technology	M1259-01/02
Mag-Bind® Endo-Free® Plasmid Mega Kit	Isolation of up to 5 mg of Endotoxin free plasmid DNA using magnetic beads	M6262-02

Bulk: For most products bulk quantities are available please inquire.

Please Call, Fax , or e-mail us to place an order.
We will be happy to answer any questions for you.

Tel: 770-931-8400 (US) Fax: 770-931-0230 (US) e-mail: info@omegabiotek.com
Tel: 1-800-832-8896 (Toll free) Fax: 1-888-624-1688 (Toll free)
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